Notes

THE ISOLATION AND STRUCTURE OF CP-120,509, A NEW POLYETHER ANTIBIOTIC RELATED TO SEMDURAMICIN AND PRODUCED BY MUTANTS OF Actinomadura roseorufa

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(Received for publication January 6, 1992)

Following the discovery of the semi-synthetic anticoccidial ionophore semduramicin^{1,2)} (UK-61,689), a mutation program was undertaken to produce this polyether antibiotic by direct fermentation³⁾. The producing culture of the antibiotic

UK-58,852, Actinomadura roseorufa ATCC 53666, was chosen as the parental strain to be mutagenically treated to induce a semduramicin-producing mutant^{4,5)}. The two compounds differ from each other in that the A-ring sugar moiety of UK-58,852 is replaced by a hydroxyl group in semduramicin. Recently, we described the formation of two new polyether antibiotics, CP-91,243 and CP-91,244, isolated from the fermentation broths of two mutants of the parental strain⁶⁾. These new antibiotics were co-produced with UK-58,852. Ultimately, the direct fermentation of semduramicin, free of UK-58,852, CP-91,243 and CP-91,244, was accomplished using other mutants of this same parental strain³⁾. Interestingly, an unknown minor metabolite was observed together with semduramicin. We now report the isolation and characterization of this minor metabolite, the novel polyether CP-120,509 (1).

The production of semduramicin by direct fermentation of *A. roseorufa* ATCC 53664 (obtained



THE JOURNAL OF ANTIBIOTICS





(2.61 g)

Table 1. Physico-chemical properties of CP-120,509 free acid (1) and Na-salt (1-Na).

Property	1	1-Na	
MP	80~82°C	143~145°C	
$[\alpha]_{\rm D}^{25}$ (c 1.0, CHCl ₃)	$+32.3^{\circ}$	$+28.3^{\circ}$	
Empirical formula	$C_{45}H_{76}O_{17}$	$C_{45}H_{75}O_{17}Na$	
MW	889.10	911.07	
Elemental analysis			
Calcd:	С 60.79, Н 8.62	C 59.33, H 8.30	
Found:	C 60.18, H 9.06	С 59.11, Н 8.87	
IR (CHCl ₃) cm ⁻¹	3360, 2940, 2900, 2845, 1725 (-CO ₂ H), 3060, 2940, 2895, 2840, 1590 (1460, 1380, 1110, 1060, 990, 970 1460, 1400, 1375, 1110, 1050 970		
Solubility			
Soluble	Organic solvents	Organic solvents	
Insoluble	H ₂ O	H ₂ O	

from the mutation of strain ATCC 53666 with 1-methyl-3-nitro-1-nitrosoguanidine) was performed as described elsewhere³⁾. Although this fermentation was free of the parent antibiotic UK-58,852, a minor metabolite (*ca.* 2% relative to semduramicin based on HPLC analysis) was detected that was more polar than semduramicin. Since we sought to determine the nature of this minor metabolite, we were fortunate to find that another mutant of ATCC 53666, designated mutant 2634-606, produced this same polar metabolite in a greater quantity relative to semduramicin (18:82 ratio, respectively, by HPLC analysis). From the isolation method shown in Scheme 1, 17.6-liters of fermentation broth using this latter mutant afforded 2.61 g of CP-120,509 Na-salt (1-Na). TLC analysis with silica gel plates using $CHCl_3$ - methanol (9:1) gave an Rf value of 0.52 for CP-120,509 and 0.56 for semduramicin. The antibiotics were visualized by spraying the TLC plate with vanillin-EtOH-

Carbon	CP-120,509		Semduramicin ^a	
	¹³ C shift ^b	¹ H shift ^e	¹³ C shift ^b	¹ H shift ^e
1 COONa	178.51 (0)	_	179.09 (0)	
2 CH_2	44.97 (2)	2.22, 2.62	45.38 (2)	2.17, 2.49
3 O-Č-O	97.81 (0)		97.70 (0)	
4 CH	45.33 (1)	1.47	45.28 (1)	1.48
5 O-CH	75.20 (1)	3.62	74.74 (1)	3.71
6 O-CH	82.42 (1)	2.98	81.95 (1)	3.11
7 O–CH	67.55 (1)	3.73	66.80(1)	3.73
8 CH	33.93 (1)	1.88	33.74 (1)	1.98
9 O-CH	68.60 (1)	3.97	67.61 (1)	4.23
10 CH	34.51 (1)	1.84	33.54 (1)	1.81
11 O-CH	69.83 (1)	3.95	70.03 (1)	3.92
12 CH ₂	33.21 (2)	1.66, 1.94	33.77 (2)	1.62, 1.90
13 O-C-O	106.80 (0)	_	107.45 (0)	_
14 CH ₂	39.16 (2)	1.74, 1.94	38.89 (2) ^d	1.73, 1.97
15 CH ₂	33.54 (2)	1.72, 1.96	33.39 (2) ^d	1.76, 1.98
16 C-O	84.41 (0)	—	84.51 (0)	—
17 O-CH	82.45 (1)	3.45	82.28 (1)	3.53
18 CH ₂	28.44 (2)	1.48, 1.67	26.83 (2)	1.47, 1.71
19 CH ₂	32.17 (2)	1.40, 2.63	32.25 (2)	1.50, 2.40
20 C–O	83.43 (0)	_	84.15 (0)	
21 O-CH	87.19(1)	3.81	86.96 (1)	4.03
22 O-CH	80.76 (1)	4.10	80.92 (1)	4.16
23 CH ₂	33.21 (2)	2.12, 2.30	32.47 (2)	2.23
24 O-CH	77.98 (1)	4.52	80.22 (1)	4.49
25 O-CH	74.76 (1)	3.83	73.01 (1)	3.93
26 CH	31.60 (1)	1.34	33.11 (1)	1.23
27 CH ₂	35.26 (2)	1.32, 1.46	36.40 (2) ^d	1.32, 1.42
28 CH	35.97 (1)	1.45	39.81 (1)	1.43
29 O-C-O	97.69 (0)		96.89 (0)	
4-Me	12.30 (3)	1.04	12.10 (3)	1.03
6-OMe	60.17 (3)	3.53	59.03 (3)	3.52
8-Me	10.76 (3)	1.07	11.05 (3)	1.08
10-Me	10.50 (3)	0.83	10.43 (3)	0.84
16-Me	27.31 (3)	1.41	27.56 (3)	1.49
20-Me	24.67 (3)	1.18	23.25 (3)	1.12
26-Me	16.69 (3)	0.82	17.51 (3)	0.87
28-Me	16.15 (3)	0.81	16.99 (3)	0.91
29-CH ₂ OH	65.01 (2)	3.23, 3.96	—	_
29-Me	_		26.05 (3)	1.29
1' 0-CH-0	103.30 (1)	4.41	103.22 (1)	4.41
2' CH2	30,53 (2)	1.53 1.77	30.55 (2)	1 53 1 80
3' CH_	26.87 (2)	1.32 2.18	26.92 (2)	1.31 2 18
4' O-CH	79.82 (1)	2.80	79.83(1)	2.81
5' O-CH	74.60 (1)	3.31	74,57 (1)	3.31
4'-OMe	56.85 (3)	3.35	56.86 (3)	3.36
5'-Me	18.38 (3)	1.26	18.38 (3)	1.24

Table 2. ¹³C and ¹H NMR chemical shift data for the Na-salts of CP-120,509 and semduramicin in CDCl₃.

^a For details of spectral assignment of semduramicin Na-salt see ref 6.

^b In ppm from TMS in CDCl₃ solution; number of attached protons are in parentheses.

[°] In ppm from TMS in CDCl₃ solution.

^d Previous assignment (ref 6) has been confirmed in a recent study of the biosynthesis of semduramicin, *i.e.*, incorporation of [1-¹³C]propionate, [1-¹³C]acetate, or [1,2-¹³C₂]acetate: J. P. DIRLAM, H. A. I. MCARTHUR & E. B. WHIPPLE, unpublished results.

e 4'-O-Methylamicetose.



 H_2SO_4 reagent followed by heating at 100°C.

The physico-chemical properties of 1 and 1-Na are given in Table 1. Spectroscopic data and elemental analysis were consistent with C45H76O17 for free acid 1, and C₄₅H₇₅O₁₇Na for sodium salt 1-Na. For example, in the low resolution positive FAB-MS, a diagnostic cationized molecule m/z 911 $(M+Na)^+$ was detected for 1-Na, where M is defined as the mass of the free acid (calcd for $C_{45}H_{76}O_{17} + Na: 911.4980$). Furthermore, 1-Na gave a base peak at m/z 849, 62 daltons less than the corresponding metal-adduct molecular ion, which is common for polyethers having a β hemiketal carboxylic acid group $((M + Na - CO_2 (H_2O)^+)^{7)}$. Thus, the molecular formula for 1-Na was assumed to contain one oxygen (16 daltons) more than that for semduramicin Na-salt (C45H75O16Na).

The ¹³C, ¹H and DEPT⁸⁾ NMR data for 1-Na, recorded on a Bruker AM-500 spectrometer, revealed the following groups: CH₃ (8), CH₂ (11), CH (5), CH₃O (2), O-CH (12), C-O (2), O-CH-O (1), O-C-O (3), and -COONa (1) (Table 2). It was apparent that the structure of 1-Na was very similar to semduramicin Na-salt, but 1-Na was missing the methyl group at $\delta_{\rm C}$ 26.05 ppm (29-Me) and contained an additional methylene group at $\delta_{\rm C}$ 65.01. From a consideration of the chemical shift of the new methylene group, hydroxymethyl substitution at C-29 seemed a likely possibility.

Using ¹³C DEPT, COSY and HETCOR experi-

ments in the manner previously described⁹⁾ for ionophore CP-84,657, the spectrum of 1-Na was systematically assigned (Table 2), except for three methylene units (C-14, C-15 and C-27) which were assigned by analogy with semduramicin Na-salt, where a detailed NMR analysis had already been performed⁶⁾. The observed shifts for the terminal hydroxymethyl at C-29 ($\delta_{\rm C}$ 65.01; $\delta_{\rm H}$ 3.23 and 3.96) are in excellent agreement with those reported for the terminal hydroxymethyl at C-25 in monensin ($\delta_{\rm C}$ 64.83; $\delta_{\rm H}$ 3.23 and 3.93)¹⁰.

The structure of 1 was later confirmed by X-ray crystallography using a single crystal of the silver salt of 1. All diffractometer data were collected at -100°C. A computer generated perspective drawing of 1-Ag (Fig. 1) clearly supports the above proposed structure of 1. Interestingly, another X-ray structure of 1-Ag was determined in the same experiment owing to a disorder of the sugar moiety. This second structure (not shown) was identical with the one shown in Fig. 1 with the exception that the substituents O1', methoxy (O4a',C4b') and methyl (C5b') on the sugar ring were all in axial positions (these substituents are all equatorial in Fig. 1). The two structures, both of which had the sugar group in a chair conformation, were present in nearly a 1:1 ratio.

A small quantity of 1-Na was isolated from the fermentation of *A. roseorufa* ATCC 53664, in which 1-Na was detected at only 2% of the semduramicin level. The NMR spectral data obtained using this

sample were in good agreement with those reported in Table 2. The high resolution positive FAB-MS showed a cationized molecule with an exact mass at m/z 911.4980 ((M+Na)⁺; calcd for C₄₅H₇₆O₁₇+ Na: 911.4980). Finally, both samples were analyzed by reversed-phase HPLC on a Partisil C₈ column (10 μ m; 4.6 × 250 mm) using RI detection; elution was by MeCN-H₂O (95:5) with a flow rate of 1.0 ml/minute. A retention time of 4.83 minutes was found for the two samples of 1-Na. Using these HPLC conditions, the following retention times were determined: CP-91,243 (4.41 minutes), semduramicin (5.13 minutes), CP-91,244 (5.66 minutes) and UK-58,852 (7.78 minutes).

CP-120,509 exhibited *in vitro* antibiotic activity against certain Gram-positive bacteria, and the spirochete, *Serpulina* (*Treponema*) *hyodysenteriae* (the causative agent of swine dysentery), but was inactive against Gram-negative bacteria. It afforded excellent anticoccidial activity against *Eimeria acervulina* in chickens at levels between 30 and 60 mg/kg in feed; however, it was devoid of activity versus *Eimeria tenella* at levels as high as 60 mg/kg in feed. In the same test, semduramicin was superior to CP-120,509 against *E. acervulina* at 15 mg/kg in feed.

Acknowledgments

The authors are grateful to Ms. D. M. RESCEK for NMR spectral data and Dr. J. G. STROH and Mr. K. J. ROSNACK for FAB-MS. We thank Mr. A. R. GAUTHIER for performing some of the HPLC analyses. We are indebted to Mr. S. B. SEIBEL for the antimicrobial assays and Dr. J. E. SHIVELY (Pfizer Animal Health, Terre Haute, IN) for conducting the anticoccidial studies.

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